

Research Article

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Low cost nanoparticles derived from nitrogen fertilizer industry waste for the remediation of copper contaminated soil and water

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Abstract

The goal of this study was to produce a novel nano-scale material from nitrogen fertilizer industry byproduct (nNFIB) and assess its capability to remediate Cu contaminated wastewater and soil. The novel nNFIB was produced using planetary mono mill and characterized. Equilibrium and kinetics studies of Cu sorption by nNFIB were performed in batch system. The effects of a variety of factors, including pH, coexisting ions and adsorption time on Cu adsorption were investigated. Furthermore, Cu sequestration mechanism onto nNFIB was investigated using sequential extraction technique and Fourier transform infrared (FTIR) spectra before and after nNFIB adsorption. The Cu sorption equilibrium and kinetics data were successfully described by Langmuir and first-order models, respectively. The calculated maximum Cu(II) adsorption capacity (qmax) of nNFIB (100 mg g–1) was four times higher than qmax of bulk NFIB. Copper removal by nNFIB was quite fast (around 86%) in the first 5 min and gradually slowed down until achieved 100% removal at equilibrium time. The FTIR spectra and Cu fractionation data in biosolid-amended soil demonstrated that Cu sequestration in contaminated water and soil is strongly related to CaCO₃ of nNFIB. The overall findings show the potential use of nNFIB as a best management practice for Cu removal from wastewater and Cu stabilization in contaminated biosolid-amended soils.

Keywords: Biosolid-amended sandy soil, Cu fractionation, Cu sequestration, FTIR spectra, Sorption and kinetic models, Wastewater



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1 **1. Introduction**

Anthropogenic activities have polluted the environment with excess amounts of heavy metals. 2 Rapid industrialization and urbanization have discharged huge amounts of heavy metals into 3 ecosystems and metals such as copper are introduced mainly by electroplating and metal mining 4 industries and water pipe work [1]. Because copper is very toxic at low concentration, copper-5 contaminated wastewater must be treated before discharging to protect the environment. 6 Exposure to excessive amounts of copper can cause serious health problems because of its bio-7 accumulation and high toxicity [2, 3]. United State Environmental Protection Agency (USEPA) 8 reported that the permissible limit of copper ions in industrial effluents is 1.3 mg/L while World 9 Health Organization (WHO) stated that copper ions content in drinking water should not exceed 10 1 mg/L [4, 5]. Therefore, the development of low-cost, yet efficient and environment-friendly 11 12 technologies to remediate Cu contaminated water and soil is urgent to protect and preserve public and ecosystem health. 13

Remediation of Cu contaminated water and soil can be performed by different technologies including chemical/electro-chemical processes, ion exchange, reverse osmosis, coagulation, and adsorption [6, 7].The removal of metal ions better accomplished by adsorption due to its safety, simplicity and the advancement of adsorption technology. However, the newly introduced adsorbents such as active carbon, grapheme oxide and mesoporous silica are economically unattractive [8].

In recent years, sustainable resources have become a greater concern and research has focused on the production of low cost sorbents from industrial byproducts such as eggshells, oyster shells, water treatment residuals, Fly ash and iron slags and the ability of these byproducts

to remove aqueous pollutants have also been tested [9-11]. There is a wealth of recent articles 1 regarding industrial and agricultural byproducts as a precursor [12]. However, information is 2 lacking on sorbents produced from calcite-based waste materials generated from nitrogen 3 4 fertilizer industry as a precursor materials and the feasibility of using it in water and soil remediation. Worldwide, millions of tons of calcite-based solid waste from nitrogen fertilizer 5 industry are generated each year. Surprisingly, no information or systematic study regarding the 6 use of NFIB for Cu²⁺ removal has been reported. Thus, the overall purpose of this work is to 7 determine the ability of calcite-based materials generated from nitrogen fertilizer industry 8 byproducts to remediate contaminated soil and water. 9

The specific objectives of the current study were to: (1) produce and characterize a novel nanomaterials derived from nNFIB and examine the feasibility of using such materials as simple yet effective adsorbents for aqueous copper removal and immobilization (2) optimize the various parameters governing Cu²⁺ removal such as pH, contact time, and coexisting ions (3) investigate Cu sequestration mechanism onto nNFIB. For the best of authors' knowledge there are no other articles in existence concerning the use of nanoparticles derived from nitrogen fertilizers industry byproducts for remediation of copper contaminated soil and water.

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18 2. Material and Methods

19 2.1. Preparation and Characterization of nNFIB

The NFIB was obtained from nitrogen fertilizer company, Alexandria, Egypt. The NFIB samples
were collected, air-dried, ground and passed through two different sieves having pore diameters
of 2 mm (mNFIB) and 51µm (µNFIB). Nanoparticles of NFIB were produced using subsamples

of μ NFIB (< 51 μ m) and Planetary Mono Mill according to the method of Elkhatib et al. [13]. 1 The crystalline nature of nNFIB was determined using Bruker AXS D8 Advance X-ray 2 Diffractometer. The particles size, morphology and elemental composition of nNFIB were 3 investigated using scanning electron microscope equipped with energy dispersive spectroscopy 4 (SEM-EDS) (INCAx-Sight, Oxford Instruments, UK). Surface area of nNFIB was determined 5 Resea using autosorb iQ surface area analyzer (Quanta chrome, USA). 6

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2.2. Sorption Isotherms and Kinetics 8

Copper (II) sorption equilibrium studies were performed on mNFIB and nNFIB at normal pH 9 10 (7.2) using 0.01M KNO₃ and Cu (II) concentrations ranging from10-500 mg/L. The NFIB-Cu mixtures (in replicate) were equilibrated on a shaker for 24 h, centrifuged for 10 min at 4000 rpm, 11 filtered through a membrane filter $(0.45\mu m)$ and the filtrate was analyzed for Cu by Atomic 12 13 absorption spectroscopy(AAS). Seven sorption isotherm models were assessed for their ability to fit the sorption data [14]. The sorbed Cu by nNFIB was examined via SEM equipped with an 14 EDS (INCAx-Sightmodel 6587, Oxford Instruments, UK). 15

Batch sorption kinetic experiments were conducted with Cu at room temperature (25 \pm 16 2°C). A known dose of nNFIB (150 mg) was mixed with 20 mL of Cu (II) solutions with initial 17 18 concentration of 500 mg/L in 50 mL centrifuge tubes. The mixtures (in replicate) were shaken for different time intervals (5 min - 24 h) using an end-over-end shaker at 3 different pH levels 19 (pH 5, 7 and 9). The pH was kept constant by automatic titration with HCl or NaOH. The Cu -20 21 nNFIB suspensions were centrifuged and then filtered using 0.45μ m Milliporefilter. Atomic absorption spectrometry (AAS, contrAA300) was used to analyze Cu (II) concentrations in the 22

supernatant solutions. Stock standard Cu (NO₃)₂ solution (1000 mgL⁻¹) was used to prepare Cu(II)
solutions. The kinetics of Cu sorption on the nNFIB samples were investigated by fitting the
sorption data to power function, parabolic diffusion, first order, and Elovich kinetic models [9].

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5 2.3. Spiking Biosolids and Incubation Experiment

Biosolids samples originally contained 12.80 mg Cu kg⁻¹ were collected and spiked with Cu 6 nitrate at a rate of 300 mg Cu kg⁻¹, the soil Cu concentration limit recommended by the USDA 7 [15]. The Cu-spiked biosolids samples were incubated under aerobic conditions at room 8 temperature $(25 \pm 2^{\circ}C)$ and 70% water holding capacity for 30 d and then were mixed with 2 kg 9 10 of sandy soil. Four different rates of nNFIB (0, 0.25, 0.5, and 1%) were added to soil -biosolids mixtures, placed in plastic bags, and incubated for 20 d at room temperature ($25 \pm 2^{\circ}$ C). During 11 the incubation period, moisture content of the mixtures was maintained at 70% of water holding 12 13 capacity (WHC).

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15 2.4. Copper Fractionation

The procedure of Tessier et al. [16] was used to fractionate Cu in soil-biosolids mixtures before and after application of nNFIB. The used procedure fractionates Cu into five fractions: Exchangeable (Exch), carbonates (Carb), Fe–Mn oxides (FeMnO), organic matter (OM) and Residual (Res). Copper in the fractions was determined using AAS, contr AA300.

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1 **2.5. Statistical Analysis**

The experimental data were statistically analyzed using Microsoft Excel and COSTAT programs 2 Fisher's least significant difference at level of significance $P \le 0.01$ was used to separate 3 esearch 4 differences among the treatment means [17].

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6 **3. Results and Discussions**

3.1. Characterization of Nanoparticles From nNFIB 7

The XRD patterns of bulk NFIB and nanoscale NFIB (Fig. S1) demonstrate a strong 8 characteristic peak at $2\theta = 30^{\circ}$ indicating that both samples are mainly containing high 9 percentage (93%) of calcite (CaCO₃). The SEM and EDX analyses of both samples confirmed 10 XRD results and ascertained that the main component of bulk and nanoscale NFIB is calcite (Fig. 11 12 1(a), (b)). The SEM image of nNFIB sample before Cu saturation (Fig. 1(b)) showed the spherical shape of nanoparticles and the representative single particle sizes(< 100 nm) in 13 diameter. The SEM image of Cu saturated nNFIB, also revealed that adsorbed Cu has formed a 14 coating layer on nNFIB surface (Fig. 1(c)) and the presence of Cu peak $(3.70\% \pm 0.76)$ has been 15 confirmed by EDX spectrum of Cu saturated nNFIB (Fig. 1(c)) .Meanwhile, EDX analysis 16 showed a decrease in calcium percentage from 93.4 to 91.3% of the total elements in nNFIB as a 17 result of Cu ions addition (Fig. 1(b), (c)). 18

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Fig. 1. Scanning electron microscopy (SEM) image and energy dispersive X-ray (EDX) elemental
distribution of (a) mNFIB, (b) nNFIB and (c) the Cu-loaded nNFIB.

1 **3.2.** Sorption Isotherm

Sorption isotherm studies were performed to determine the maximum sorption capacity for the 2 three different particle sizes of NFIB (m NFIB, µ NFIB and n NFIB). Copper sorption isotherms 3 (Fig. 2(a)) have shown continuous increase of sorbed Cu by nNFIB, uNFIB and mNFIB with 4 increasing Cu concentration from 5 to 500 mg L⁻¹. The Cu adsorption capacity of nNFIB was 5 higher than that of uNFIB and mNFIB. Reliable prediction of Cu adsorption parameters 6 including maximum sorption capacity was further analyzed using 7 sorption isotherms models 7 [13, 18] (Table 1). Langmuir and Temkin models best described Cu sorption data in the order: 8 Langmuir > Temkin as evidenced by the high determination coefficient (R^2) and low standard 9 error of estimate (SE) values of these models (Table 1). 10

The superiority of Langmuir model to describe the adsorption behavior of Cu(II) on 11 nNFIB, uNFIB and mNFIB (Fig. 2(b), Table 1) indicates the involvement of monolayer 12 adsorption in the Cu(II) removal process by nNFIB,uNFIB and mNFIB. Based on Langmuir 13 model, the calculated maximum Cu(II) adsorption capacity (q_{max}) of nNFIB is 100 mg g⁻¹ which 14 is four times higher than that of Bulk NFIB. This is not surprising since the BET specific surface 15 area of nNFIB (225.4 m² g⁻¹) is much higher than that of the bulk NFIB sample (8.8 m² g⁻¹). 16 17 High surface area of nanoparticles greatly enhances surface reactivity and the adsorption capacity of nNFIB [17]. Therefore, producing NFIB in nanoscale will greatly enhance its 18 capability to remove Cu (II) from contaminated wastewater. 19

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Fig. 2. (a) Copper sorption isotherm onto mNFIB, uNFIB and n NFIB materials. (b) Langmuir
 sorption isotherms.

Table 1. Equilibrium Model Constants and Standard Error of Estimate (SE) and Determination Coefficients (R^2) for Copper Adsorption by the Three Different Particles Sizes of NFIB

Models	Parameter	m NFIB	μ NFIB	n NFIB
Freundlich	K _F (mL g ⁻¹)	5.009×10 ³	5.519×10 ³	8.119 ×10 ³
$q_e = K_F C_e^{1/n}$				
	1/n	0.255	0.294	0.413
	R ²	0.49	0.62	0.78
	SE	0.98	0.88	0.78
Langmuir	q _{max} (µgg⁻)	25000	30000	100000
$q_e = q_{max}(K_L C_e / 1 + K_L C_e)$	K _L (L mg ⁻¹)	0.25	0.29	0.33
	R ²	0.99	0.96	0.94
	SE	7.4×10 ⁻⁵	6.69 ×10 ⁻⁵	1.36 ×10⁻⁵
Elovich	q _{max} (µgg ¹)	1.67×10^{4}	2.00 ×10 ⁴	2.50 ×10 ⁴
$q_e/q_{max} = K_E C_e \exp(-qe/qmax)$	K _E (L mg ⁻¹)	1.647	1.135	1.341
	R ²	0.25	0.49	0.50
	SE	0.463	0.329	0.412
Temkin	$\Delta Q(k \text{ J mol}^{-1})$	6.275	9.044	26.313
$\theta = RT/\Delta Q \ln K_0 C_e$	$K (1 \sigma^{-1})$	20 227	24.062	20 020
		0.09	24.002	20.030
	SE	0.56	0.57	0.051
Fowler-Guggenheim(FG)	$W(k mol^{-1})$	-3 221	-0.680	5 134
Fowler=Guggermenn(rG)		5.221	0.000	3.134
$K_{FG}C_e = 0/1 - 0 \exp(20 \text{ W/RT})$	K _{FG} (L mg ⁻¹)	1.467	2.228	0.718
	R ²	0.94	0.80	0.97
	SE	0.221	0.109	0.348
Kiselev	$k_1(L mg^{-1})$	1.476	1.449	0.489
$k_1 C_e = \theta / (1 - \theta) (1 + k_n \theta)$	kn	1 913	1 034	2 570
	R ²	0.80	0.95	0.97
	SE	5.122	3.012	2.679
Hill-deBoer	$K_1(Lmg^{-1})$	1.270	1.077	0.498
$K = 0 / (1 - 0) \exp(0 / (1 - 0) - K - 0/RT)$				
$\kappa_1 c_e = 0$ (1 = 0) exp(0) (1 = 0) = $\kappa_2 0$ (1)	K_2 (kJ mol ⁻¹)	12.164	12.717	3.704
	R ²	0.99	0.98	0.94
	SE	0.088	0.249	0.132

 $\begin{array}{ll} q_e \ (mg \ g^{-1}) \ is \ Cu \ adsorbed \ per \ gram \ of \ adsorbent, \ C_e \ (mg \ L^{-1}) \ is \ equilibrium \ Cu \ concentration \ in \ solution, \ K_f \ is \ a \ constant \ related \ to \ adsorption \ capacity \ of \ the \ adsorbent \ (Lmg^{-1}), \ n \ is \ a \ constant, \ q_{max} \ (mg \ g^{-1}) \ is \ the \ maximum \ adsorption \ capacity \ of \ the \ adsorbent, \ K_L \ (Lmg^{-1}), \ is \ Langmuir \ constant \ related \ to \ the \ free \ energy \ of \ adsorption, \ \theta \ is \ fractional \ coverage, \ R \ is \ the \ universal \ gas \ constant \ (klmol^{-1} \ K^{-1}), \ T \ is \ the \ temperature \ (K), \ \Delta Q \ is \ (-\Delta H) \ the \ variation \ of \ adsorption \ energy \ (klmol^{-1}), \ K_0 \ is \ Temkin \ constant \ (Lmg^{-1}), \ K_{FG} \ is \ Fowler-Guggenheim \ constant \ (Lmg^{-1}), \ W \ is \ the \ interaction \ energy \ between \ adsorbed \ molecules \ (klmol^{-1}), \ K_1 \ is \ Kiselev \ constant \ (Lmg^{-1}), \ K_n \ is \ a \ constant \ of \ complex \ formation \ between \ adsorbed \ molecules, \ K_1 \ is \ Hill-de \ Boer \ constant \ (Lmg^{-1}), \ K_1 \ is \ Constant \ related \ to \ the \ interaction \ between \ adsorbed \ molecules. \ A \ positive \ K_2 \ means \ attraction \ between \ adsorbed \ species \ and \ a \ negative \ value \ means \ repulsion. \$

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9 **3.3. Effect of Adsorbent-Sorbate Contact Time**

The effects of contact time (5 min to 24 h) on Cu (II) adsorption by nNFIB at three different pH 10 values (5, 7, and 9) are presented in Fig. 3(a). Over 86% of Cu (II) was removed by nNFIB in the 11 first 5 min and then slowed down to reach 100% removal at the end of the 24 h (equilibrium 12 time). The fast Cu removal is attributed to the high proportion of calcium carbonate (more 93%) 13 in nNFIB and with time the availability of adsorption active sites rendered non available. 14 Increasing the initial pH of aqueous solutions from 5 to 9 increased the removal efficiency of Cu 15 (II) by nNFIB (Fig. 3). With increasing pH values (pH > 7), the surface charges of nNFIB 16 became more negative and that may cause a greater ion-exchange reaction between Cu(II) and 17 Ca(II) and greater sequestration of Cu(II) at the nNFIB surface [19]. These results are also 18 19 supported by other researchers [20, 21].

Copper adsorption data at three pH values (5, 7, and 9) were kinetically analyzed and fitted to Elovich, first-order, Parabolic diffusion, and power function models [22, 23] (Table S1). The power function and first-order kinetics models best described copper (II) adsorption on nNFIB in the order: power function model > first-order as evidenced by their highest R^2 and lowest SE values (Table S1, Fig. 3). The adsorption rate (K_a) of the power function model increased from 5.71 × 10⁴ to 6. 70 × 10⁴ min⁻¹ with the increase in the system pH from 5 to 9 (Table S1) which indicates that Cu sorption is preferably at high pH values.



2 3.4. Mechanism of Copper (II) Removal by nNFIB

The FTIR Spectra of nNFIB was generated in the frequency range 350 - 4400 cm⁻¹ before and 3 after copper adsorption to elucidate Cu (II) adsorption mechanism (Fig. 4). The broad band at 4 3451 cm⁻¹ in the FTIR spectrum of nNFIB is assigned to the OH stretching vibrations of the 5 hydroxyl molecule [24]. The vibration bands at 1439 cm⁻¹, 875 cm⁻¹, 712 cm⁻¹ and 409 cm⁻¹ 6 indicate plane bending vibration of carbonate [25-29]. After Cu(II) adsorption by nNFIB, the 7 band at 3451 cm⁻¹ was decreased in the intensity and shifted to lower wave number (3437 cm⁻¹). 8 In addition, the intensities of the bands corresponded to amorphous calcium carbonate at 875 and 9 712 cm⁻¹ clearly increased and the CO₃⁻¹ symmetric stretch at 1398 cm⁻¹ shifted to 1439cm⁻¹. 10 These shifts are typical for Cu²⁺complexation by OH group and precipitation as copper(II) 11 carbonate. Therefore, FTIR analysis demonstrated the involvement of OH and CO₃⁻⁻ groups in 12 13 Cu retention by nNFIB and suggested that Cu (II) adsorption by nNFIB at high pH (> 7) may take place via OH bonding colloidal insoluble hydroxides, Cu (OH)₂ and surface precipitation as 14 copper(II) carbonate. 15

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17 3.5. Copper Removal in Single and Multi-Element System by nNFIB

The effect of coexisting ions on Cu removal by nNFIB was studied by using two competing cations (Cd and Pb) at concentrations equal to Cu concentration. Copper removal by nNFIB was markedly affected by the presence of Cd (II) and Pb (II) at160 mgL⁻¹ concentration (Fig. 5). The Cu removal efficiency of nNFIB decreased by 17.6% (from 98.6% to 81.0%), 5.0% (from 98.8% to 93.8), 6.1% (from 99.2% to 93.1%), and 4.9% (from 99.5% to 94.6) at Cu concentrations 5, 20,



1 3.6. Effect of nNFIB Application on Cu (II) Sequestration in Contaminated Soil

The effect of nNFIB application on distribution of Cu fractions in contaminated biosolids 2 amended soil is presented in Fig. 6. The percentages of copper fractions in nNFIB unamended 3 4 soil followed the order: Res > FeMnO > OM > Exch > Carb. Application of nNFIB to the contaminated biosolids amended soil at rates of 0.25, 0.50 and 1.0% greatly reduced the Exch-Cu 5 and Res fractions and simultaneously increased carbonate fractions. In the soil amended with 1.0% 6 nNFIB, ~ 37% of Cu (II) was associated with the carbonate fraction, whereas organic (13.66%), 7 and exchangeable (6.46%) fractions represent the minor association. nNFIB application 8 significantly increased Cu association with the carbonate fraction and consequently enhanced Cu 9 10 (II) immobilization in the soil studied. Therefore, it is suggested that the use of nNFIB for Cu sequestration could geochemically stabilize heavy metals in contaminated soils. 11

To further investigate Cu sequestration mechanism onto nNFIB treated soil, FTIR spectra 12 13 of biosolids amended sandy soil before and after nNFIB application were performed. The FTIR spectrum of the biosolids amended sandy soil (Fig. 7) displays prominent bands at 3406, 1435, 14 876 and 1033cm⁻¹ which are attributed to OH bonded water, stretching and vibration modes of 15 16 carbonate groups, and Si-O-Si bending vibrations, respectively [20, 28, 34]. Application of 17 nNFIB to biosolids amended sandy soil has resulted in a shift of OH bonded water band at 3406cm⁻¹ to a higher wave number (3419 cm⁻¹) and a reduction of its intensity due to the strong 18 interaction between OH groups and Cu ions. Meanwhile, the intensity of the band attributed to 19 calcite at 1433 cm⁻¹ greatly increased as a result of increasing CO_3^{2-} content in the nNFIB 20 21 biosolids amended sandy soil sample due to nNFIB application (Fig. 7). The FTIR results have

sorption by nNFIB [35-37]. Exchangeable Carbonat Fe-Mn oxides OM RS earch Cu fraction (%) 0% 0.25% 0.5% 1% nNFIB nNFIB nNFIB nNFIB Fig. 6. Percentage of Cu fractions in biosolids amended soil after nNFIB application at rates of 0, 0.25, 0.5, and 1% by weight. Cu loaded-andy-Biosolid-nNFIE Cu loaded-Sandy-Biosolid Transmittance (%) Sandy-Biosolid Sandy Soil 500 350 Wavenumber (Cm⁻¹)

clearly shown that OH and CO3²⁻ groups are the main functional groups responsible for Cu (II)

Fig. 7. Fourier transmission infrared (FTIR) spectra of Cu-loaded sandy soil and amended with biosolids
before and after nNFIB application.

1 4. Conclusions

2 A novel nano-structured adsorbent (nNFIB) derived from the low cost waste of nitrogen fertilizer industry was developed. The high surface area of nanoparticles greatly enhanced surface 3 4 reactivity and Cu(II) adsorption capacity of nNFIB. The capability of NFIB nanoparticles to remove Cu(II) reached 4 times higher than that of bulk NFIB. First-order kinetics model was 5 best suited to the kinetics data. Copper adsorption was affected by the pH of solution and the 6 background ions. FTIR results demonstrated the important rules of hydroxyl and carbonate 7 groups in sequestering Cu (II) from contaminated soils and water. In brief, it can be stated that 8 nNFIB is a promising adsorbent to remove Cu from wastewater and to stabilize Cu in 9 contaminated biosolid-amended soils. 10

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14

15 Author Contributions

16 Elsayed Elkhatib (Professor) provided the intellectual input and the protocols to be followed in

17 the study, wrote the manuscript and lead the overall study (corresponding author).

Mohamed Moharem (Associate Professor) carried out Cu analyses and assist in calculations,drawings and interpretation.

20 Ayman Mahmoud (Master) conducted all the experiments.

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